

SAMPLING PROTOCOL FOR PARASITE KIT

Sampling Apparatus

- The sampling apparatus does not have to be sterile but it must be clean and uncontaminated by cysts and/or oocysts.
- Thoroughly clean the apparatus, including filter holder, hoses and pumps, and rinse between samples. If multiple samples are to be collected with the same apparatus, arrange sampling sequence to begin with the least contaminated water (eg., treated drinking water) and end with the most contaminated water (eg., source water).
- If field conditions preclude to a thorough cleaning of apparatus components between samples, thoroughly rinse all surfaces that will come in contact with the water with at least 50 gal of water to be sampled prior to the installation of the filter cartridge.

Sample Collection

- Use a water-resistant marking pen to record the start time, volume filtered, date, and sampling location onto bag.
- Water flow should be 3 to 4 GPM, rate is read a widest point of float.
- A minimum sample size of at least 100 gal is required.
- After the required volume of water has passed through the filter, shut off the water flow, record the stop time. Disconnect sampling apparatus while maintaining the inlet hose level above the level of the opening on the outlet hose in order of prevent backwashing and the loss of particulate matter from the filter.
- Aseptically remove the filter from the holder and transfer to labeled bag. Seal the bag and place it inside a second bag.
- Transport the sample to the laboratory on wet ice or cold packs and refrigerated at 2-5°C.

Do not freeze during transport or storage

- Sample should be submitted to the laboratory within 96 hrs.

PROPOSED ICR PROTOZOAN METHOD FOR DETECTING GIARDIA CYSTS & CRYPTOSPORIDIUM OOCYSTS IN WATER
BY A FLUORESCENT ANTIBODY PROCEDURE
ICR Revised: Oct 29, 1993

Location: 2062 W. Henry Ave.
Sidney, B.C. V8L 5Y1
Tel: (250)656-1334
Fax: (250)656-0443

Mailing: P.O. Box 2103
Sidney, B.C. V8L 3S6

Email: mblabs@pacificcoast.net